

Case Study

# Optimization of the kinetics of siRNA desorption from the surface of silicon nanoparticles

Anzhelika Melnikova<sup>1,2\*</sup>, Roman Kirkin<sup>1</sup> and Luidmila Komarova<sup>1</sup>

<sup>1</sup>Obninsk Institute for Nuclear Power Engineering, Obninsk, Russia

<sup>2</sup>A. Tsyb Medical Radiological Research Centre, Russia

## Abstract

Oncological diseases are one of the most significant medical and social diseases in most countries of the world. Over the past decades, the search and development of new drugs, treatment regimens and methods of molecular diagnostics of malignant neoplasms remains relevant. In turn, an important goal of molecular genetic research is to suppress the expression of genes responsible for the development of tumors. The key targets taken into account in the development of antitumor drugs are proteins involved in carcinogenic changes in the cell. One of the promising molecular targets for the development of medicinal compounds in targeted therapy of tumor diseases is poly(ADP-ribose)polymerase 1 (PARP1). A potential way to inhibit PARP1 even at the stage of protein translation is RNA interference due to small interfering RNAs (siRNAs). For the penetration of siRNAs into the target cell, it is necessary to develop a method of their transportation controlled in space and time. An actual direction for solving this problem is the use of highly stable porous silicon-based nanoparticles. In the current study, in order to increase the functionality of nanoparticles, their surface was modified with various agents (functionalization), providing increased efficiency of drug loading and more uniform release.

## Introduction

One of the promising molecular targets for the development of medicinal compounds in targeted therapy of tumor diseases is poly(ADP-ribose)polymerase 1 (PARP1) [1,2]. The gene of the same name encoding this protein has increased expression in a number of oncological diseases, therefore it is considered an important molecular target in the development of antitumor agents.

Poly(ADP-ribosyl) action is a reaction that occurs when a cell responds to genotoxic stress. This process, being one of the posttranslational modifications of nuclear proteins, occurs in the presence of enzymes of the PARP poly(ADP-ribose) polymerase family, which play the role of recognizing breaks. The activity of members of the PARP family (PARP1 and PARP2) is associated with cellular signaling pathways. During poly(ADP-ribosyl)ation they ultimately contribute to changes in gene expression, the amount of RNA and proteins, as well as the localization and activity of proteins that mediate signaling responses. It is also crucial for a wide range of physiological and pathological responses and, thus, is

a good target for chemical therapy of a number of diseases [3]. Preclinical data suggest that PARP inhibitors can enhance the effects of radiation therapy in certain types of tumors, namely: lung, colorectal cancer, head, and neck cancer, glioma, cervical cancer and prostate.

To disrupt the repair process in tumor cells, it is necessary to take into account the mechanisms of action of PARP inhibitors. PARP1 controls several DNA repair pathways: excision repair of bases (base excision repair, BER) and nucleotides (NER), mismatch repair (MMR), repair of double-stranded breaks using homologous recombination (HR) and non-homologous end joining (NHEJ) [4,5].

The most popular are biochemical methods of inhibiting PARP 1, such as enzymes and other agents. However, a potential way to reduce the influence of this factor even at the stage of protein translation is RNA interference.

RNA interference is the process of suppressing gene expression, which is carried out by small interfering RNAs (siRNAs). These are short RNAs that bind to the matrix RNA, thereby inhibiting the translation process [6].

### More Information

#### \*Address for correspondence:

Anzhelika Melnikova, Obninsk Institute for Nuclear Power Engineering, Obninsk, Russia, Email: angelik\_melnikova@mail.ru

Submitted: February 17, 2023

Approved: March 01, 2023

Published: March 02, 2023

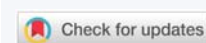
**How to cite this article:** Melnikova A, Kirkin R, Komarova L. Optimization of the kinetics of siRNA desorption from the surface of silicon nanoparticles. *Ann Biomed Sci Eng.* 2023; 7: 020-023.

DOI: 10.29328/journal.abse.1001021

 <https://orcid.org/0000-0001-7229-2813>

**Copyright license:** © 2023 Melnikova A, et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**Keywords:** Silicon nanoparticles; PARP1; RNA interference; siRNA



For the penetration of siRNAs into the target cell, it is necessary to develop a method of their transportation controlled in space and time. A promising direction for solving this problem is the use of highly stable porous silicon-based nanoparticles (NPS) [7-16].

The issue of the genotoxicity of nano derivatives remains insufficiently investigated, and the methodology for assessing their genotoxic effects and predicting the genotoxic risk of human use has not yet been developed. The assessment of the genotoxicity of nanoparticles is fragmentary and contradictory and does not provide a prediction of the genotoxic risks of exposure of nanomaterials to humans.

Silicon nanoparticles belong to ceramic nanomaterials, along with aluminum and titanium nanoparticles. Most often they are made of porous material and are very common as carriers of medicines during antitumor therapy. The vast majority of silicon nanoparticles, including those protected by one or another coating, release more than 90% of therapeutic RNAs within 10-15 hours. The functionalization of polyethyleneimine (PEI) and (2-aminoethyl)-3-aminopropyltrimethoxysilane (DEMO-P) nanoparticles allows direct interaction with cancer cells and effective penetration into its microenvironment compared to traditional delivery systems [17].

The aim of our work was to optimize silicon nanoparticles for delivering small interfering RNAs against the PARP1 gene and ensuring their controlled desorption.

## Materials and methods

To conduct the siRNA loading study, we used mesoporous NPS with a size of 120 nm. The study used siRNAs against one of the key transcription factors (PARP1). Nanoparticles were obtained by laser ablation and fragmentation in liquid. Laser ablation in liquid makes it possible to synthesize new ultrapure nanomaterials (without side contamination), which can make a decisive contribution to biomedical applications. In order to increase the functionality of the NPS, their surface was modified with various agents (functionalization), providing increased efficiency in loading the drug and more uniform release.

The experiment used untreated nanoparticles, as well as nanoparticles functionalized with polyethyleneimine (PEI) and (2-aminoethyl)-3-aminopropyltrimethoxysilane (DAMO-P) to study the possibility of loading RNA.

After that, the siRNA was loaded into the pores of the nanoparticles (in the case of untreated NPS) or onto their surface due to chemical bonds (in the case of functionalized NPS). For untreated nanoparticles, as well as for nanoparticles functionalized by PEI, there were 4 loading modes with ultrasound duration of 30, 60, 120 and 240 seconds.

The most important indicator for evaluating the

effectiveness of nanoparticles as a means of transporting drugs is the desorption kinetics, that is, the rate of release of the drug (in the specific case of siRNA) from nanoparticles or from their surface.

To identify the kinetics of siRNA desorption from the surface of silicon nanoparticles, a regular concentration measurement was carried out. Measurements were performed on a NanoDrop ND-1000 spectrophotometer. Immediately before the experiments, suspensions of nanoparticles were subjected to ultrasonic treatment to eliminate the agglomerates formed during storage. The concentration of nanoparticles in suspensions was measured by the gravimetric method: by drying a certain volume of suspensions with an unknown concentration and measuring the mass of the dried substance.

Statistical analysis of data was performed using RStudio for Windows and Microsoft Excel. The nonparametric Chi-Square criterion is used to assess the reliability of the results [18].

## Results

The study showed that silicon nanoparticles without additional modification (pSi) demonstrate an extremely low level of siRNA loading (about 1%), which is due to the retention of siRNA solely due to silicon pores. Loading siRNA onto silicon nanoparticles functionalized with PEI-pSi showed significantly higher efficiency (up to 10%). This is explained by the chemical binding of siRNA and the imino group in PEI. The insufficiently high loading rate is due to the small amount of PEI on the pSi surface.

The most optimal duration of ultrasonic treatment of a solution of nanoparticles and siRNA is 60 seconds. During this time, additional dispersion of nanoparticles occurs, and RNA almost does not degrade. This treatment provides the longest and most uniform yield of siRNA.

The functionalization of nanoparticles with 2-aminoethyl-3-aminopropyl-trimethoxysilane (DAMO-pSi) makes it possible to load more than 90% of siRNA during ultrasonic processing, which is almost 10 times more efficient than when processing PEI. Loading without ultrasound demonstrates significantly lower efficiency, which is explained by the aggregation of nanoparticles. When using DAMO-pSi, a sharp desorption intensity is observed (within 24 hours), which is less effective for RNA interference.

## Discussion

Loading siRNA onto silicon nanoparticles functionalized with polyethyleneimine (PEIpSi) showed significantly higher efficiency (up to 10%). This is explained by the chemical binding of siRNAs and specifically the groups within PEI. The insufficiently high loading rate is due to the small amount of PEI on the pSi surface. The functionalization of nanoparticles with 2-aminoethyl-3-aminopropyl-triethoxysilane (DEMO-



pSi) allows loading more than 90% of siRNA during ultrasonic processing, which is almost 10 times more efficient than when processing PEI. Loading without ultrasound demonstrates significantly lower efficiency, which is explained by the aggregation of nanoparticles. When using DEMO-pSi, a sharp desorption intensity is observed (within 24 hours), which is less effective for RNA interference.

## Conclusion

Small interfering RNAs have a great potential for wide application in biomedicine, in particular, in the treatment of malignant diseases [19]. PARP1, a key DNA repair protein, and the most important transcription factor is overexpressed in tumor cells, and therefore is a promising target for RNA interference. In the course of the study, the design of the siRNA sequence against PARP1, which does not affect other genes, was carried out.

For the transport of siRNA, nanoparticles of porous silicon are used as a biocompatible and biodegradable material. Various methods of surface coating and functionalization are used to absorb siRNA nanoparticles by target cells. The nature of the coating also determines the kinetics of the release of siRNA from nanoparticles [20].

Functionalization of silicon nanoparticles with polyethyleneimine demonstrates a low degree of loading but is a good indicator of desorption. When treated with silane, the loading efficiency is much higher, but there is a sharp release of siRNA. The optimal ultrasound treatment time is 60 seconds. The use of untreated nanoparticles is ineffective due to the low level of loading.

The results obtained indicate that the delivery of siRNA to target cells using treated silicon nanoparticles is a promising method for the treatment of oncological diseases. However, to confirm the effectiveness of such delivery, it is necessary to refine the methods of nanoparticle functionalization and RNA loading, as well as the use of molecular methods to determine the level of gene expression.

## Funding

This research was supported by the Obninsk Institute of Atomic Energy.

## References

- Ludwig A, Behnke B, Holtlund J, Hiltz H. Immunoquantitation and size determination of intrinsic poly(ADP-ribose) polymerase from acid precipitates. An analysis of the in vivo status in mammalian species and in lower eukaryotes. *J Biol Chem.* 1988 May 25;263(15):6993-9. PMID: 3130376.
- Yamanaka H, Penning CA, Willis EH, Wasson DB, Carson DA. Characterization of human poly(ADP-ribose) polymerase with autoantibodies. *J Biol Chem.* 1988 Mar 15;263(8):3879-83. PMID: 3126180.
- Gibson BA, Kraus WL. New insights into the molecular and cellular functions of poly(ADP-ribose) and PARPs. *Nat Rev Mol Cell Biol.* 2012 Jun 20;13(7):411-24. doi: 10.1038/nrm3376. PMID: 22713970.
- De Lorenzo SB, Patel AG, Hurley RM, Kaufmann SH. The Elephant and the Blind Men: Making Sense of PARP Inhibitors in Homologous Recombination Deficient Tumor Cells. *Front Oncol.* 2013 Sep 11;3:228. doi: 10.3389/fonc.2013.00228. PMID: 24062981; PMCID: PMC3769628.
- Thomas C, Tulin AV. Poly-ADP-ribose polymerase: machinery for nuclear processes. *Mol Aspects Med.* 2013 Dec;34(6):1124-37. doi: 10.1016/j.mam.2013.04.001. Epub 2013 Apr 25. PMID: 23624145; PMCID: PMC3750069.
- Hammond SM, Bernstein E, Beach D, Hannon GJ. An RNA-directed nuclelease mediates post-transcriptional gene silencing in *Drosophila* cells. *Nature.* 2000 Mar 16;404(6775):293-6. doi: 10.1038/35005107. PMID: 10749213.
- Rao W, Wang H, Han J, Zhao S, Dumbleton J, Agarwal P, Zhang W, Zhao G, Yu J, Zynger DL, Lu X, He X. Chitosan-Decorated Doxorubicin-Encapsulated Nanoparticle Targets and Eliminates Tumor Reinitiating Cancer Stem-like Cells. *ACS Nano.* 2015 Jun 23;9(6):5725-40. doi: 10.1021/nn506928p. Epub 2015 May 29. PMID: 26004286.
- Min Y, Caster JM, Eblan MJ, Wang AZ. Clinical Translation of Nanomedicine. *Chem Rev.* 2015 Oct 14;115(19):11147-90. doi: 10.1021/acs.chemrev.5b00116. Epub 2015 Jun 19. PMID: 26088284; PMCID: PMC4607605.
- Joo J, Kwon EJ, Kang J, Skalak M, Anglin EJ, Mann AP, Ruoslahti E, Bhatia SN, Sailor MJ. Porous silicon-graphene oxide core-shell nanoparticles for targeted delivery of siRNA to the injured brain. *Nanoscale Horiz.* 2016 Sep 1;1(5):407-414. doi: 10.1039/C6NH00082G. Epub 2016 Jun 14. PMID: 29732165; PMCID: PMC5935492.
- Gowda R, Jones NR, Banerjee S, Robertson GP. Use of Nanotechnology to Develop Multi-Drug Inhibitors For Cancer Therapy. *J Nanomed Nanotechnol.* 2013 Dec;4(6):184. doi: 10.4172/2157-7439.1000184. PMID: 25013742; PMCID: PMC4085796.
- Cheng YJ, Luo GF, Zhu JY, Xu XD, Zeng X, Cheng DB, Li YM, Wu Y, Zhang XZ, Zhuo RX, He F. Enzyme-induced and tumor-targeted drug delivery system based on multifunctional mesoporous silica nanoparticles. *ACS Appl Mater Interfaces.* 2015 May 6;7(17):9078-87. doi: 10.1021/acsami.5b00752. Epub 2015 Apr 24. PMID: 25893819.
- Lerner MI, Glazkova EA, Lozhkomoiev AS, Svarovskaya NV, Bakina OV, Pervikov AV, Psakhie SG. Synthesis of Al nanoparticles and Al/AlN composite nanoparticles by electrical explosion of aluminum wires in argon and nitrogen. *Powder Technology.* 2016; 295: 307–314.
- Ramani M, Ponnusamy S, Muthamizhchelvan C, Marsili E. Amino acid-mediated synthesis of zinc oxide nanostructures and evaluation of their facet-dependent antimicrobial activity. *Colloids Surf B Biointerfaces.* 2014 May 1;117:233-9. doi: 10.1016/j.colsurfb.2014.02.017. Epub 2014 Mar 4. PMID: 24657608.
- Alarifi S, Ali D, Alkahtani S. Nanoalumina induces apoptosis by impairing antioxidant enzyme systems in human hepatocarcinoma cells. *Int J Nanomedicine.* 2015 May 25;10:3751-60. doi: 10.2147/IJN.S82050. PMID: 26045665; PMCID: PMC4448921.
- Mikhaylov G, Mikac U, Magaeva AA, Itin VI, Naiden EP, Psakhie I, Babes L, Reinheckel T, Peters C, Zeiser R, Bogyo M, Turk V, Psakhie SG, Turk B, Vasiljeva O. Ferri-liposomes as an MRI-visible drug-delivery system for targeting tumours and their microenvironment. *Nat Nanotechnol.* 2011 Aug 7;6(9):594-602. doi: 10.1038/nnano.2011.112. PMID: 21822252.
- Zhang S, Li J, Lykotrafitis G, Bao G, Suresh S. Size-Dependent Endocytosis of Nanoparticles. *Adv Mater.* 2009;21:419-424. doi: 10.1002/adma.200801393. PMID: 19606281; PMCID: PMC2709876.
- Zhang M, Xu R, Xia X, Yang Y, Gu J, Qin G, Liu X, Ferrari M, Shen H. Polycation-functionalized nanoporous silicon particles for gene silencing on breast cancer cells. *Biomaterials.* 2014 Jan;35(1):423-31. doi: 10.1016/j.biomaterials.2013.09.033. Epub 2013 Oct 5. PMID: 24103653; PMCID: PMC3842233.



18. Korovin MS, Bakina OV, Fomenko AN, Lerner MI. Assessment of cytotoxic effect of low-dimensional aluminum oxide structures on tumor cells. *Siberian journal of oncology*. 2016; 15(6): 35-41.
19. Wan Y, Apostolou S, Dronov R, Kuss B, Voelcker NH. Cancer-targeting siRNA delivery from porous silicon nanoparticles. *Nanomedicine (Lond)*. 2014 Oct;9(15):2309-21. doi: 10.2217/nnm.14.12. Epub 2014 Mar 5. PMID: 24593001.
20. Tong WY, Alnakhli M, Bhardwaj R, Apostolou S, Sinha S, Fraser C, Kuchel T, Kuss B, Voelcker NH. Delivery of siRNA in vitro and in vivo using PEI-capped porous silicon nanoparticles to silence MRP1 and inhibit proliferation in glioblastoma. *J Nanobiotechnology*. 2018 Apr 13;16(1):38. doi: 10.1186/s12951-018-0365-y. PMID: 29653579; PMCID: PMC5898074.